

Potential anti-inflammatory treatments for chronic wounds

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Abstract

Skin is the primary protective barrier against physical, chemical, thermal and infectious threats in the environment. Maintaining the integrity of skin is a matter of survival. The collaborative term – wound healing – describes the complex, yet well-organised, network of biological processes that enables tissue injuries to resolve. The many stages include haemostasis, inflammation, matrix synthesis, angiogenesis, re-epithelialisation, contraction and remodelling; often simplified to inflammation, tissue proliferation and maturation. Advances in our understanding of the influence of immune cells, and growth factors and cytokines at each stage of wound healing have led to the development of potential treatments. Here we review the biology of chronic wound healing and discuss the potential of epicatechin gallate and activated protein C to promote wound healing in a clinical setting.

Keywords: wound healing, leg ulcer, activated protein C, epicatechin gallate.

Introduction

Pressure, venous and diabetic leg ulcers are the most common type of chronic wounds affecting the Australian population. The prevalence of chronic leg ulcers in the community is 0.11%, with 24% of ulcerations persisting for more than one year¹. As a result, 45% of ulcer patients suffer immobility problems, with many being house-bound. Despite wound healing techniques dating back to the Edwin Smith papyrus in 1700 BC, treatments for chronic wounds have largely fallen short. While there has been a movement from 'dry dressings' to 'growth factors' and 'silver treatment' these treatments still struggle to obtain and/or maintain wound closure, and cost the Australian Government an excessive amount in wound management costs². More comprehensive anti-inflammatory treatments that promote angiogenesis and re-epithelialisation may be more advantageous. In this review we discuss the biology of chronic wound healing and potential new treatments in wound management, epicatechin gallate (ECG) and activated protein C (APC).

Biology of chronic wound healing

Foremost in treating chronic wounds is to understand their pathobiology. The interplay of immune cells, growth factors and cytokines during the various stages of wound healing is key to the success of wound resolution. Injury to blood vessels leads to the extravasation of blood constituents and the formation of a clot. As one of the first cell types activated after injury, platelets secrete a number of growth factors such as epidermal growth factor (EGF), platelet-derived growth factor (PDGF), and cytokines including transforming growth factor- β (TGF- β). In addition, disruption of the epidermal barrier promotes secretion of interleukin (IL)-1

and tumour necrosis factor- α (TNF- α) by keratinocytes. These chemical signals attract inflammatory cells to the site of injury. Neutrophils and monocytes are recruited to the site of injury where they clear the wound site of contaminating bacteria and debris. After debridement, macrophages release inflammatory cytokines including IL-1, IL-6, EGF, PDGF, TGF- β , fibroblast growth factor (FGF) and vascular endothelial growth factor (VEGF) to stimulate the formation of granulation tissue and neovascularisation by fibroblasts and endothelial cells respectively. In chronic wounds, a number of factors contribute to impaired healing, including:

1. Prolonged inflammation

In normal wound healing, neutrophils are cleared from the wound after 72 hours by macrophages. Exudates collected by lavage from the surface of ulcers have shown that 95% of inflammatory cells are neutrophils³. Macrophages are infrequent and inactive, and CD4⁺ T-lymphocytes are evident at the wound edge⁴. The persistence of inflammatory cells in the wound space contributes to elevated levels of cytokines, including IL-1 α , IL-1 β , IL-6 and TNF- α ⁵, which promote the inflammatory environment and stimulate the production of tissue-degrading proteinases⁶.

2. Delayed granulation tissue formation

Prior to wound closure the wound space must be filled by granulation tissue – a combination of extracellular matrix (ECM) proteins including fibronectin, vitronectin and subsequently type I collagen. In chronic wounds, these ECM proteins are increasingly degraded by elevated levels of inflammatory cell-derived proteinases, including urokinase, cathepsin-G, neutrophil-derived elastase, and

matrix metalloproteinases (MMP)-1, MMP-2, MMP-8, MMP-9⁷⁻⁹. These proteinases also degrade cytokines and growth factors required by fibroblasts for the deposition of ECM proteins (PDGF, TGF- β and FGF)^{5,10}. At the same time, there is a relative decrease in proteinase inhibitors such as α 1-antitrypsin, α 2-macroglobulin and tissue inhibitor of metalloproteinases-1 due to inactivation^{11,12}. These inhibitors normally act to regulate the activity of proteinases in chronic wound fluid. The resultant imbalance of proteinases and inhibitors, and reduction in fibroblast growth factors, leads to a delay in granulation tissue deposition, and subsequent new blood vessel formation (or neovascularisation) and re-epithelialisation of the wound.

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3. Delayed neovascularisation and re-epithelialisation

Akin to fibroblasts, endothelial cells and keratinocytes are dependent on growth factors and cytokines for neovascularisation and re-epithelialisation of the wound, respectively. Therefore, the above-mentioned proteinase-mediated degradation of growth factors and cytokines (PDGF, TGF- α , IGF-I, FGF and EGF) in chronic wounds contributes to delayed wound healing^{5,10}.

An important premise in chronic wound therapy is to tailor treatment to the aetiology of the wound whenever possible. According to factors that contribute to impaired healing, the ideal treatment would control inflammation to enable progression through to the proliferative stages; granulation tissue formation, neovascularisation and re-epithelialisation. Deficiency of neutrophils and macrophages using anti-neutrophil serum and *PU.1* null mice has shown that the removal of inflammatory cells from a wound is crucial for accelerated wound healing and reduced scar formation^{13,14}. Two agents that have potential to meet these criteria are ECG and APC.

Epicatechin gallate (ECG)

Catechins are a family of polyphenolic flavonoids found in green tea extracts. The three major isoforms, epigallocatechin gallate (EGCG), ECG and epicatechin, have anti-oxidant, -proliferative, -tumour and -inflammatory properties¹⁵. While *in vitro* studies assessing the mechanisms of ECGs actions are not clearly understood, *in vivo* studies have demonstrated a potential role for ECG in chronic wound healing.

Anti-inflammatory

Catechins decrease inflammatory cell infiltration¹⁶ and production of pro-inflammatory cytokines IL-1 β , IL-6 and TNF- α , while enhancing the production of the anti-inflammatory cytokine, IL-10 by human inflammatory cells^{17,18}. The suppression of inflammatory cytokines by catechins is mediated by reduced nuclear factor *kappa* B (NF- κ B) activation¹⁸.

Granulation tissue deposition, neovascularisation and re-epithelialisation

ECG inhibits MMP-9 and MMP-2 activity¹⁹, and collagen degradation by binding to collagen and preventing tissue-degrading enzymes binding²⁰. Subcutaneous injection of ECG at the wound margin enhances collagen deposition and maturation, increases vascular endothelial growth factor and accelerates angiogenesis²¹. ECG also increases the activity of enzymes, inducible nitric oxide synthase and cyclooxygenase-2, required for the re-epithelialisation of

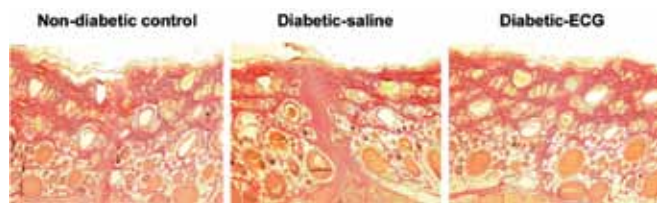


Figure 1. Representative micrographs of wound healing and scar formation of incisional wounds in non-diabetic control, and diabetic mice treated with saline or ECG at 14 days. Subcutaneous injections of saline or ECG were given daily for seven days. Magnification 3X.

wounds²¹. In the full-thickness wound healing rat model, ECG increases granulation tissue formation, neovascularisation and re-epithelialisation of wounds.

Wound healing

In a murine model of type II diabetes, ECG accelerated wound healing²². Figure 1 shows the quality of wound healing and scar formation at 14 days in non-diabetic control, and diabetic wounds treated with saline or 0.8 mg/ml ECG daily for seven days. Diabetic wounds treated with saline demonstrated a wide, clearly visible wound tract with high proportions of disorientated immature (pink) collagen fibres indicating poor wound healing. By contrast, non-diabetic and diabetic wounds treated with ECG exhibited near complete wound contracture and a high proportion of mature (red) collagen fibres orientated parallel to the epidermis, comparable to the surrounding skin. These results indicate that ECG may improve the healing of diabetic ulcers. Similar results of improved cellular organisation of granulation tissue, neovascularisation, and re-epithelialisation in another murine model of type II diabetes have been demonstrated using EGCG²³.

Activated protein C (APC)

Activated protein C (APC) is an anticoagulant with anti-inflammatory properties. The PC zymogen is activated to APC by serine proteases after which it can engage the anti-inflammatory receptors endothelial protein C receptor (EPCR) and protease-activated receptor (PAR)-1^{24,25}. Acting via these receptors, APC has demonstrated a number of wound healing actions.

Anti-inflammatory

In vitro and *in vivo* studies have revealed that APC has strong anti-inflammatory properties associated with a reduction in inflammatory cells and inflammatory cytokines. For example, APC suppresses neutrophil migration by binding to the $\beta 1$ and $\beta 3$ integrin adhesion molecules²⁶, and inhibits the activation of NF- κ B in various cell types including keratinocytes²⁷. The NF- κ B pathway is important for the production of

inflammatory cytokines, including TNF- α and cell adhesion molecules associated with inflammation. Specific treatment of monocytes with APC decreases the production of the pro-inflammatory cytokines TNF- α , IL-1 β , IL-6, and IL-8^{28,29}, and increases anti-inflammatory cytokine IL-10 production³⁰.

Granulation tissue deposition, neovascularisation and re-epithelialisation

Treatment of cultured human endothelial cells or keratinocytes with APC inhibits apoptosis and promotes cell proliferation^{31,32}, migration and survival³³, necessary for neovascularisation and re-epithelialisation of the wound. Studies using the chick chorioallantoic membrane assay have shown that APC not only stimulates angiogenesis but causes epithelial cells to grow across the top of the gelatin sponge, mimicking keratinocyte re-epithelialisation in *in vivo* wound healing³⁴. APC differentially regulates proteinase activity by suppressing MMP-9 activity³⁵, a collagen-degrading MMP associated with inflammatory conditions, yet increases anti-inflammatory MMP-2 activity in keratinocytes^{34,36}, endothelial cells³⁶ and dermal fibroblasts³⁷.

Wound healing

The extensive *in vitro* research on APC and its potential mechanisms for wound healing led to its application in animal models, and subsequently pilot human studies^{34,38}. Rodent models of wound healing showed that a single topical application of 20 μ g APC reduced neutrophil infiltration, increased neovascularisation and significantly reduced the wound size of full-thickness punch biopsy wounds by day 4, with no adverse effects³⁴. In an open pilot study of four patients with lower leg ulcers of varied cause, topical application of APC once a week for four weeks stimulated the formation of granulation tissue and re-epithelialisation with no adverse effects or safety issues reported³⁸.

Figure 2 shows an example of an APC-treated patient whose deep chronic ulcer on his heel had been present for four years with no response to standard wound treatment. Striking features of healing following APC application in this example

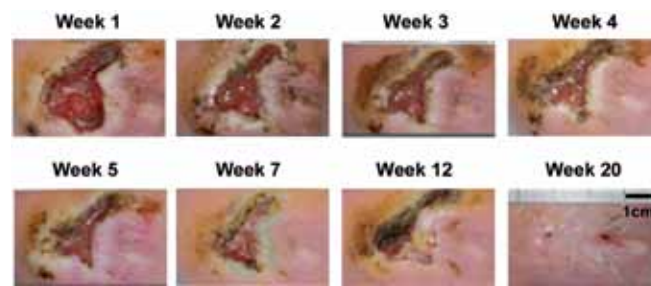


Figure 2. Progress of a patient with a four-year-old wound on the base of his heel. Topical APC treatment was given weekly for six weeks. Scale bar 1 cm.

were rapid improvement after a single treatment (week 2), and the formation of refractile translucent matrix (weeks 4–7) and granulation tissue to fill in the wound from its base. These results provide evidence that APC has the potential to improve healing in chronic wounds. Similar results were obtained in a study investigating topical APC treatment in conjunction with topical negative pressure, to treat long-standing orthopaedic wounds³⁹.

Future directions

Ideal chronic wound treatments would inhibit excessive inflammation, whilst stimulating keratinocyte and endothelial cell proliferation, migration and survival. Both ECG and APC appear to have the ability to stimulate keratinocyte and endothelial cell growth whilst inhibiting inflammation. The therapeutic potential of ECG or APC, either alone or combined, as a topical treatment for chronic wounds requires confirmation in double blind, placebo-controlled randomised clinical trials.

References

- Baker SR & Stacey MC. Epidemiology of chronic leg ulcers in Australia. *ANZ J Surg* 1994; 64:258–61.
- MacLellan D. Chronic leg ulceration—the hidden epidemic. *Med J Aust* 1994; 161(10):619–21.
- Diegelmann RF. Excessive neutrophils characterize chronic pressure ulcers. *Wound Repair Regen* 2003; 11(6):490–95.
- Moore K, Ruge F & Harding K. T lymphocytes and the lack of activated macrophages in wound margin biopsies from chronic leg ulcers. *Br J Dermatol* 1997; 137(2):188–94.
- Mast BA & Schultz GS. Interactions of cytokines, growth factors, and proteases in acute and chronic wounds. *Wound Repair Regen* 1996; 4(4):411–20.
- Postlethwaite A *et al.* Interleukin 1 stimulation of collagenase production by cultured fibroblasts. *J Exp Med* 1983; 157(2):801–6.
- Wysocki AB, Staiano-Coico L & Grinnell F. Wound Fluid from Chronic Leg Ulcers Contains Elevated Levels of Metalloproteinases MMP-2 and MMP-9. *J Invest Dermatol* 1993; 101(1):64–8.
- Rogers AA *et al.* Involvement of proteolytic enzymes—plasminogen activators and matrix metalloproteinases—in the pathophysiology of pressure ulcers. *Wound Repair Regen* 1995; 3(3):273–83.
- Nwomeh BC *et al.* MMP-8 Is the Predominant Collagenase in Healing Wounds and Nonhealing Ulcers. *J Surg Res* 1999; 81(2):189–95.
- Yager DR *et al.* Ability of chronic wound fluids to degrade peptide growth factors is associated with increased levels of elastase activity and diminished levels of proteinase inhibitors. *Wound Repair Regen* 1997; 5(1):23–32.
- Grinnell F, Ho C-T & Wysocki A. Degradation of fibronectin and vitronectin in chronic wound fluid: analysis by cell blotting, immunoblotting, and cell adhesion assays. *J Invest Dermatol* 1992; 98(4):410–6.
- Grinnell F & Zhu M. Fibronectin degradation in chronic wounds depends on the relative levels of elastase, alpha1-proteinase inhibitor, and alpha2-macroglobulin. *J Invest Dermatol* 1996; 106(2):335–41.
- Simpson D & Ross R. The neutrophilic leukocyte in wound repair: A study with antineutrophilic serum. *J Clin Invest* 1972; 51(8):2009–23.
- Martin P *et al.* Wound Healing in the PU.1 Null Mouse—Tissue Repair Is Not Dependent on Inflammatory Cells. *Curr Biol* 2003; 13(13):1122–8.
- Chan MM-Y *et al.* Inhibition of Inducible Nitric Oxide Synthase Gene Expression and Enzyme Activity by Epigallocatechin Gallate, a Natural Product from Green Tea. *Biochem Pharmacol* 1997; 54(12):1281–6.
- Maruyama T *et al.* Supplementation of green tea catechins in dentifrices suppresses gingival oxidative stress and periodontal inflammation. *Arch Oral Biol* 2011; 56(1):48–53.
- Crouvezier S *et al.* The effects of phenolic components of tea on the production of pro- and anti-inflammatory cytokines by human leukocytes *in vitro*. *Cytokine* 2001; 13(5):280–6.
- Yang F *et al.* Green Tea Polyphenols Block Endotoxin-Induced Tumor Necrosis Factor-Production and Lethality in a Murine Model. *J Nutr* 1998; 128(12):2334–40.
- Isemura M *et al.* Inhibition of Matrix Metalloproteinases by Tea Catechins and Related Polyphenols. *Ann N Y Acad Sci* 1999; 878(1):629–31.
- Jackson J *et al.* The inhibition of collagenase induced degradation of collagen by the galloyl-containing polyphenols tannic acid, epigallocatechin gallate and epicatechin gallate. *J Mater Sci Mater Med* 2010; 21(5):1435–43.
- Kapoor M *et al.* Effects of Epicatechin Gallate on Wound Healing and Scar Formation in a Full Thickness Incisional Wound Healing Model in Rats. *Am J Pathol* 2004; 165(1):299–307.
- McKelvey K & Appleton I. Epicatechin gallate improves healing and reduces scar formation of incisional wounds in type 2 diabetes mellitus rat model. *Wounds* 2012; 24(3):55–7.
- Kim H *et al.* Enhanced wound healing by an epigallocatechin gallate-incorporated collagen sponge in diabetic mice. *Wound Repair Regen* 2008; 16(5):714–20.
- Riewald M *et al.* Activated protein C signals through the thrombin receptor PAR1 in endothelial cells. *J Endotoxin Res* 2003; 9(5):317–21.
- Riewald M *et al.* Activation of endothelial cell protease activated receptor 1 by the protein C pathway. *Science* 2002; 296(5574):1880–2.
- Elphick GF *et al.* Recombinant human activated protein C inhibits integrin-mediated neutrophil migration. *Blood* 2009; 113(17):4078–85.
- Xue M *et al.* Activated protein C stimulates proliferation, migration and wound closure, inhibits apoptosis and upregulates MMP-2 activity in cultured human keratinocytes. *Exp Cell Res* 2004; 299(1):119–27.
- Grey ST *et al.* Selective inhibitory effects of the anticoagulant activated protein C on the responses of human mononuclear phagocytes to LPS, IFN-gamma, or phorbol ester. *J Immunol* 1994; 153(8):3664–72.
- Stephenson DA *et al.* Modulation of monocyte function by activated protein C, a natural anticoagulant. *J Immunol* 2006; 177(4):2115–22.
- Tolt LJ *et al.* Protective effects of activated protein C in sepsis. *Thromb Haemost* 2008; 100(4):582–92.
- Uchiba M *et al.* Activated protein C induces endothelial cell proliferation by mitogen-activated protein kinase activation *in vitro* and angiogenesis *in vivo*. *Circ Res* 2004; 95(1):34–41.
- Xue M *et al.* Endothelial protein C receptor and protease-activated receptor-1 mediate induction of a wound-healing phenotype in human keratinocytes by activated protein C. *J Invest Dermatol* 2005; 125(6):1279–85.
- Jackson CJ & Xue M. Activated protein C—An anticoagulant that does more than stop clots. *Int J Biochem Cell Biol* 2008; 40(12):2692–97.
- Jackson CJ *et al.* Activated protein C prevents inflammation yet stimulates angiogenesis to promote cutaneous wound healing. *Wound Repair Regen* 2005; 13(3):284–94.
- Xue M *et al.* Differential regulation of matrix metalloproteinase 2 and matrix metalloproteinase 9 by activated protein C: relevance to inflammation in rheumatoid arthritis. *Arthritis Rheum* 2007; 56(9):2864–74.
- Nguyen M, Arkell J & Jackson CJ. Activated protein C directly activates human endothelial gelatinase A. *J Biol Chem* 2000; 275(13):9095–8.
- Xue M *et al.* Activated protein C stimulates expression of angiogenic factors in human skin cells, angiogenesis in the chick embryo and cutaneous wound healing in rodents. *Clin Hemorheol Microcirc* 2006; 34(1–2):153–61.
- Whitmont K *et al.* Treatment of chronic leg ulcers with topical activated protein C. *Arch Dermatol* 2008; 144(11):1479–83.
- Wijewardena A *et al.* Combination of activated protein C and topical negative pressure rapidly regenerates granulation tissue over exposed bone to heal recalcitrant orthopedic wounds. *Int J Low Extrem Wounds* 2011; 10(3):146–51.